

Surface-Engineered Magnetic Nanoparticle Platforms for Cancer Imaging and Therapy

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CONSPECTUS



E normous efforts have been made toward the translation of nanotechnology into medical practice, including cancer management. Generally the applications have fallen into two categories: diagnosis and therapy. Because the targets are often the same, the development of separate approaches can miss opportunities to improve efficiency and effectiveness.

The unique physical properties of nanomaterials enable them to serve as the basis for superior imaging probes to locate and report cancerous lesions and as vehicles to deliver therapeutics preferentially to those lesions. These technologies for probes and vehicles have converged in the current efforts to develop nanotheranostics, nanoplatforms with both imaging and therapeutic functionalities. These new multimodal platforms are highly versatile and valuable components of the emerging trend toward personalized medicine, which emphasizes tailoring treatments to the biology of individual patients to optimize outcomes. The close coupling of imaging and treatment within a theranostic agent and the data about the evolving course of an illness that these agents provide can facilitate informed decisions about modifications to treatment.

Magnetic nanoparticles, especially superparamagnetic iron oxide nanoparticles (IONPs), have long been studied as contrast agents for magnetic resonance imaging (MRI). Owing to recent progress in synthesis and surface modification, many new avenues have opened for this class of biomaterials. Such nanoparticles are not merely tiny magnetic crystals, but potential platforms with large surface-to-volume ratios. By taking advantage of the well-developed surface chemistry of these materials, researchers can load a wide range of functionalities, such as targeting, imaging and therapeutic features, onto their surfaces. This versatility makes magnetic nanoparticles excellent scaffolds for the construction of theranostic agents, and many efforts have been launched toward this goal.

In this Account, we introduce the surface engineering techniques that we and others have developed, with an emphasis on how these techniques affect the role of nanoparticles as imaging or therapeutic agents. We and others have developed a set of chemical methods to prepare magnetic nanoparticles that possess accurate sizes, shapes, compositions, magnetizations, relaxivities, and surface charges. These features, in turn, can be harnessed to adjust the toxicity and stability of the nanoparticles and, further, to load functionalities, via various mechanisms, onto the nanoparticle surfaces.

1. Introduction

Magnetic nanoparticles are an important class of biomaterials and have been made into various functional agents, such as agents for applications in imaging, cell labeling, drug delivery, gene delivery, and hyperthermia.¹ These previous studies have established the foundations for current efforts to construct magnetic nanoparticle-based nanotheranostic agents.²⁻⁶ Nanotheranostics embraces the conventional notion of marriage between therapeutics and diagnostics, but on the foundation of a nanoscale platform. Such an emerging technique adds another piece to the mosaic of personalized medicine and has attracted much attention in the community. The attractiveness of magnetic nanoparticles as building blocks of theranostics is at least 2-fold. First is their prequalification as MR imaging probes. Superparamagnetic iron oxide nanoparticles (IONPs), which show high magnetization in an external magnetic field but none when the magnetic field is removed, have been the most prominent T_2/T_2^* probes for magnetic resonance imaging (MRI); manganese- and gadolinium-containing particles, on the other hand, are at various stages of development, with the hope that they might replace metal-chelator complexes in a new generation of T_1 contrast agents. Second is a set of well-developed surface chemistry. This includes the capacity to fine-tune the physical parameters of a nanoparticle, such as its size, shape, crystallinity, and magnetism.¹ More importantly, this suggests the potential for postsynthetically replacing or modifying the coating materials and, in doing so, tailoring the nanoparticle's surface charge, chemical groups, and overall size.⁷

In this Account, we will introduce our work on engineering the surface of magnetic nanoparticles to enhance the nanoparticles' roles as tumor imaging and therapeutic agents. It is our hope that this may help to accelerate further progress in this promising field.

2. Basics of Nanoparticle Surface Engineering

2.1. Surface Coating and Particle Preparation. The synthesis and surface engineering of nanoparticles are closely related. Taking IONPs for instance, a classical paradigm is to coprecipitate Fe(II) and Fe(III) in a basic solution, in the presence of a polymer. The polymer then tangles with the growing nanocrystals, protecting them from overgrowth and aggregation.¹ Feridex, Combidex, and Resovist are products of this paradigm that have been marketed or are in clinical trials. Although these formulas are all coated with dextran (or its derivatives), other hydrophilic polymers have been found to be able to substitute for dextran as the coating

material. For example, we have used polyaspartic acid (PASP) to replace dextran as the reaction precursor.⁸ PASP bears both carboxyl and amino residues. It is believed that the multiple carboxylates function mainly by passivating the growing nanoparticle surface, while leaving free amine groups available for conjugation.

One drawback associated with the coprecipitation method is the suboptimal crystallinity of the products, a limitation that is partially associated with the low reaction temperature. To address this issue, there has been a trend toward replacing the coprecipitation method with a pyrolysis (or thermal decomposition)-based means, where an organic solvent with a high boiling point is used as the reaction medium. For instance, we prepared IONPs from DMF using polyvinylpyrrolidone (PVP) as the coating material.^{9,10} The resulting 8-10 nm PVP-IONPs had a magnetization of 110 emu/g of Fe, compared with 70 emu/g of Fe for Feridex.¹⁰ Solvents such as 1-octadecene and benzyl ether, which have even higher boiling points of around 300 °C, are now commonly used as the reaction media. To be compatible with such a change in solvent, the Fe precursors have been changed to compatible analogs, such as $Fe(CO)_5$, Fe(acac)₃, or Fe(oleate)₃, and the coatings have been changed to such materials as oleic acid and oleylamine. The resulting products can provide r₂ relaxivities as high as $300 \text{ mM}^{-1} \text{ s}^{-1}$, almost triple that of Feridex (about 100 mM⁻¹ s⁻¹).¹¹ It is worth noting that an improved crystallinity is not the only basis for this increase in magnetization. Rather, the size effect also plays an important role. At the nanoscale, the magnetization of particles increases with the particle size, due to the surface spin canting effect.¹² Unlike the conventional dextran-coated formulas, which have wide core size distributions, the pyrolysis-based preparation can yield products (with accurate size control¹²) up to 50 nm in diameter.

The coating materials (oleic acid and oleylamine), while proven to be better "sculptors" than dextrans, are hydrophobic. As a consequence, many types of nanoparticles made from the pyrolysis methods are not water-soluble and thus are unsuitable for bioapplications. To address this issue, many surface engineering techniques have been developed to impart water solubility (as well as various functionalities) to the nanoparticle surface.¹ These aims can be achieved, for instance, through the addition of a second, amphiphilic coating layer. Such a ligand can use its hydrophobic section to interact with the oleic acid/oleylamine layer to get anchored on the particle surface. Meanwhile, its hydrophilic section will be exposed to the



FIGURE 1. IONPs coated with (a) a triblock copolymer and (b) dopamine-plus-HSA to confer water solubility and functional extendibility.

surrounding water molecules, affording physiological stability and conjugation-friendly groups (such as amines, carboxyls and thiols). For instance, we have tried to alkylate poly(ethylenimine) (PEI)¹³ or a triblock copolymer (Figure 1a)¹⁴ with various lengths of hydrophobic chains. The resulting amphiphilic polymers can self-assemble onto a lipophilic IONP surface and confer water solubility. An alternative approach is to use a ligand that has high affinity toward the IONP surface. When mixed, it can take the place of the original oleic acid/oleylamine coating and lead to hydrophilicity. One representative class of this kind is dopamine and its analogs. With the two adjacent hydroxyl groups, dopamine (or its derivatives) can chelate with the surface Fe on IONPs and, as a consequence, replace the original coating.¹¹ To improve stability, it is common to preconjugate dopamine with a hydrophilic tail, such as poly(ethylene glycol) (PEG).

Alternatively, we have found that proteins, such as human serum albumin (HSA), can be electrostatically adsorbed onto the dopamine–IONP surface to endow the particles with water stability (Figure 1b).¹¹ Slightly different from the previously mentioned strategies, this approach can be described as two-step engineering. In the first step, we replace the original coating with dopamine in a DMSO/ CHCl₃ mixed solvent. In the second step, we add the dopamine-coated IONPs in DMSO into an HSA aqueous solution to induce the second coating. Introducing a coating layer via physical adsorption in this way is another common strategy in the surface modification of magnetic nanoparticles. It can be further extended to impart multiple coating layers with alternating charges onto a nanoparticle, the so-called

layer-by-layer (LbL) self-assembly approach,¹⁵ and is not limited to IONP modification. For instance, we successfully coupled gadopentetic acid (Gd-DTPA) with PEI (Gd-DTPA– PEI) and coated the conjugate onto silica nanoparticles.¹⁶

2.2. Surface Coating and Functionality. The development of favorable pharmacokinetics is an essential criterion in the design of nanoparticles intended for intravenous injection. We and others have identified several factors, including size, charge, and hydrophilicity, that can be selected to improve performance. Previously, magnetic nanoparticles, especially IONPs, were used primarily as contrast probes in magnetic resonance (MR) for reticuloendothelial system (RES) imaging. Instead of targeting RES organs, a more advanced avenue is to introduce targeting motifs, either protein-, peptide-, or aptamer-based, onto magnetic nanoparticles to create target-specific agents. In most cases, tethering of motifs is achieved through a bioconjugation technique, which uses mediators (such as EDC/NHS) or crosslinkers (such as N-succinimidyl-4-maleimidobutyrate) to form a covalent linkage between the two moieties.^{2,7} Surface engineering again plays a critical role in providing conjugation-friendly chemical groups, such as carboxyl, amine, or thiol, on the particle surface. To make efficient coupling and to avoid cross-linking, it is sometimes necessary to precovert chemical groups of one side. For instance, we coupled c(RGDyK), a tumor targeting motif, onto amineterminated, copolymer-coated IONPs. c(RGDyK) affords one amine group from lysine for coupling, so it is possible to use a homodimer linker, such as bissulfosuccinimidyl suberate (BS3), to achieve the coupling. However, such a measure will inevitably cause cross-linking among the same species.



FIGURE 2. (a) TEM bright field image of mPEG-*b*-PCL/MONP micelles; (b) AFM height image of alkyl-PEl2k–IONPs; (c) hysteresis loops of the MONPcontaining micelles measured at 300 K (inset shows a zoomed-in plot between -2 kOe and 2 kOe magnetic field); (d) T_2 relaxation rates $(1/T_2, s^{-1})$ of alkyl-PEl2k–IONP nanocomposites as a function of iron concentration (mM) for different polymer/SPIO ratios at (\bigcirc 0.6, (\blacksquare) 1.2, and (\diamondsuit) 2.5.

A better plan is to thiolate c(RGDyK) with agents such as *N*-succinimidyl *S*-acetylthioacetate (SATA).^{14,17} This converts the problem to a conjugation between thiol and amine, which can be achieved via the use of a heterodimer cross-linker such as *N*-succinimidyl-4-maleimidobutyrate.

Not all the function loading needs covalent conjugation. The loading of therapeutics, for instance, is often achieved through noncovalent interactions to ensure an easier release. For instance, it is a common strategy to coat nanoparticles with polycation materials and to use the conjugates as gene delivery vehicles. Due to electrostatic interaction, negatively charged RNA or DNA therapeutics are then able to be loaded onto the nanoparticle surface and to cross cell membranes, whose lipid bilayer cores are otherwise considered impermeable to polar molecules. Later on, in the endosomes/lysosomes, where the pH is lower, the RNA/ DNA cargos are released due to the proton sponge effect. Similarly, it is desirable to be able to load other small molecule-based therapeutics onto nanoparticle surfaces through physical interaction.

Overall, a set of chemistry has been developed that allows functionality loading to be accomplished in a fast, economic, and mild fashion. The means of choice is largely dependent on the chemical structure of the to-be-loaded motifs. However, in general, covalent conjugation is more utilized in the imaging setting, noncovalent loading is more seen in drug loading, and chelation chemistry is largely used in immobilization of radioisotopes.

2.3. Surface Engineering and MRI Contrast. The impact of surface engineering is not limited to imparting water solubility to nanoparticles; rather, it is also an important factor in modulation of particle r_1/r_2 relaxations. For instance, it was reported that coating thickness can affect protons' physical exclusion from magnetic field and residence time within the coating zone and, therefore, modulate particles' r_2 relaxivities.^{18,19} More prominently, the aggregates of nanoparticles (Figure 2) were found to be able to induce more efficient T_2 shortening, a feature that has been harnessed to construct nanoclusters of higher r_2 values (Figure 2).^{13,20,21} Taking alkyl-PEI2k-IONPs as an example,²² under a magnetic field of 3 T, single-IONPcontaining micelles have an r_2 relaxivity of 84 Fe mM⁻¹ s^{-1} , while multiple-IONP-containing micelles have an r_2 of up to 345 Fe mM⁻¹ s⁻¹. Such alkyl-PEI–IONPs also can be self-assembled onto any micro/nanotemplate pairing with polyelectrolytes and the anchoring density or the interparticle distance of IONP per template can be controlled by varying the coating conditions such as ionic strength. SiO₂



FIGURE 3. Alkyl-PEl2k–IONP nanocomposites adsorbed on polyelectrolyte-covered SiO_2 nanotemplates with (a) higher and (b) lower anchoring density. (Scale bar = 100 nm).

nanotemplates covered with higher IONP density (Figure 3a) displayed a 70% increase in T_2 relaxivity compared with the lower density ones (Figure 3b) and about 2.5 times higher than single alkyl-PEI2k–IONPs.¹⁵

Surface engineering has also proven useful in determining nanoparticle T_1 relaxivities, although via different mechanisms. Unlike T_2 probes, T_1 probes need to have direct contact with the surrounding water molecules to affect the proton relaxation times. The organic coating of the nanoparticles that lies between the two interfaces inevitably interferes with such an interaction. Although it has been reported that PEGylated phospholipid could coat onto pyrolysis-yielded MnO nanoparticles (MONPs) to transform them to T_1 contrast agents,²³ a potential concern is the hydrophobic zone that surrounds the MONP cores. Such a zone could disallow efficient water exchange and, as a consequence, lead to suboptimal T_1 contrast. Indeed, when we switched to particles with the dopamine-plus-HSA coating, we observed a 5-fold increase in r_1 (Figure 4).²⁴

3. Surface-Engineered Magnetic Nanoparticles for MR Imaging

As mentioned above, magnetic nanoparticles currently play an active role as probes in MRI. This includes conventional RES-targeting probes, which, after injection, are largely sequestered by immune cells, such as macrophages. While still widely utilized, such an approach has the limitation of only being applicable to RES organs, such as liver, spleen, bone marrow, and lymph nodes. There is a growing interest in using surface engineering to develop tumor-targeting nanoparticulate probes that can reach tumors, either primary or metastatic, in a wider range of organs. This can be achieved via the enhanced permeability and retention (EPR) effect, which refers to the increases in endothelial leakage and reductions in lymphatic drainage within tumors that can lead to accumulations of macromolecules or nanoparticles. Alternatively, the magnetic nanoparticles can be engineered to display surface biovectors, whose cognate receptors



FIGURE 4. (a) Phantom studies with HSA and phospholipid-coated MONPs at the same concentrations. Due to existence of a hydrophobic coating zone between the particle surface and surrounding water molecules, phospholipid coated MONPs tend to have a less prominent T_1 reducing effect. (b) r_1 relaxivity evaluation from the results of panel a.

are (1) aberrantly expressed in tumors and (2) able to serve as target biomarkers to achieve localized probe accumulation. $^{25-27}$

3.1. RES Targeting. By studying the relationships between surface properties and in vivo behaviors, one can elucidate laws that determine a particle's in vivo fate and, as a result, guide the future design of nanoformulas. For instance, we have prepared a series of PVP-IONPs with different hydrodynamic sizes.9 Both in vitro and in vivo studies have confirmed a size effect on the particles' RES sequestration. In particular, we have identified one formula, PVP-IO-37 (core size of 37 nm and hydrodynamic size of 100 nm), with a particularly prominent macrophage uptake rate. When injected systematically in a murine orthotropic Huh7 hepatocarcinoma model, we observed, at 1 h postinjection, a contrast change (Δ CNR) of 94% \pm 6% with PVP–IO-37, compared with 81% \pm 8% with Feridex (Figure 5). In another study, we tested Mn-doped iron-oxide (Mn-IO) nanoclusters as contrast probes for liver imaging.²⁰ The hydrophobic Mn-IO nanoparticles were synthesized in organic phase and then transferred into water with the help of a block copolymer mPEG-b-polycaprolactone (PCL). These Mn-IO nanoparticles self-assembled into small clusters inside micelles with a mean diameter of approximately 80 nm and an r_2 relaxivity of 270 mM⁻¹(Mn+Fe) s⁻¹. These nanoclusters induced significant contrast in the liver, resulting in a decrease in signal intensity of 80% within 5 min postinjection.



FIGURE 5. In vivo MR imaging with (a) normal mice and (b) an orthotropic Huh7 hepatocarcinoma model after injection with PVP–IO-37 and Feridex. Arrow points to tumor.

3.2. Tumor Targeting. The HSA-coated IONPs discussed above are good examples of achieving tumor targeting via passive means. Such a nanostructure can stay long in the circulation, yet extravasate significantly at tumor sites.²⁸ MRI T_2 maps of a U87MG xenograft murine model showed a drop of $29.9\% \pm 4.2\%$ in the signal intensity from the tumor area 18 h p.i.¹¹ Although the main mechanism of tumorhoming was attributed to the EPR effect, the HSA sheath is believed to have played a role, via its interaction with glycoprotein (gp60) receptor (albondin) or SPARC (secreted protein acid and rich in cysteine), in promoting the particle extravasation and tumor internalization. Likewise, HSA-coated MONPs were found to be able to accumulate in tumor. Also, in the U87MG xenograft model, signal intensity increases of 5.3% \pm 0.6%, 13.8% \pm 2.0% and 9.7% \pm 2.1% at 1, 4, and 24 h p.i. were observed on the T_1 -weighted maps.²⁴

On the other hand, we have sought ways to conjugate targeting motifs, such as RGD, onto IONPs to create smart probes.^{29,30} For instance, we have coupled RGD onto both triblock copolymer¹⁴ and PASP-coated NPs⁸ (TPIO and PASP–IO) and studied the tumor targeting and contrast capabilities of the conjugates. We observed, in both cases, a significant increase in affinity toward integrin $\alpha_V\beta_3$. This was attributable to the presence of multiple RGD peptides on a single nanoparticle surface, the so-called multivalent

effect.^{25,31} In both cases, we observed strong hypointensities in MRI images in the tumor areas after injection in a U87MG xenograft model. Postmortal immunohistological studies confirmed that the accumulation of nanoparticles in tumor was mostly mediated through RGD–integrin interaction. Notably, we found that, although many of the particles were able to extravasate and become bound to tumor cells, a large portion of the particles remained trapped in the blood vessel lumen.^{8,14} This was because, in such a model, the upregulation of integrin occurs on both tumor cells and tumor endothelial cell surfaces.^{32,33}

In most nanoparticle formulas, a great portion of the overall size is contributed by the coating materials. As we mentioned above, the coating materials are intended (1) to stabilize the nanoparticles in the physiological environment and (2) to afford a platform for functionality docking. However, it is generally believed that a smaller nanoparticle size is associated with a greater extravasation and with less immune sequestration. Therefore, it would be advantageous if we could remove the thick polymer coating layer and shift its nanoparticle-suspending job to, for instance, the added functional motifs, which fortunately are typically hydrophilic molecules. In one such effort, we conjugated c(RGDyK), via a Mannich reaction, onto catechol-coated IONPs³⁴ (Figure 6). The resulting nanoparticles had a core



FIGURE 6. (a) Conjugation of RGD onto 4-methylcatechol-coated IONP surface. (b) High-resolution TEM images of the IONPs. (c–e) MR images taken after IONP injection on a U87MG xenograft model: (c) without NPs, (d) with c(RGDyK)–IONPs, and (e) with c(RGDyK)–IONPs and a blocking dosage of c(RGDyK). (f,g) Prussian blue staining on tumor tissue samples from panels d and e. Arrow points to tumor.

size of about 4.5 nm and could be directly conjugated with c(RGDyK). Such an RGD layer plays a dual role in the nanosystem: (1) it enables integrin to bind to the nanoconjugates, and (2) it confers water solubility to the nanoconjugates. Owing to the lack of a thick polymer coating, such an RGD-conjugated IONP formula has an overall size of only \sim 8.4 nm, one of the smallest among its category.

3.3. Cell Labeling. Aside from uses as systemically injected probes, magnetic nanoparticles are also used as cell labeling reagents. This application is driven by the emergence of cell-based therapeutics and, associated with that, the need to understand the fate of exogenous cells in hosts.^{35,36} The idea is to load a sufficient amount of nanoparticles into the cells prior to their administration into the host. Surface engineering again can play a central role in determining the particles' internalization rate and toxicity.^{36,37}

A common strategy is to coat the IONP surface with polycation materials to induce endocytosis-mediated particle uptake.^{38,39} PEI analogs, especially those with long lengths and branched structures, have been widely utilized. However, high cytotoxicity has been associated with these polymers, which limits the safe incubation dose and, as a result, the cell loading rate. To address this issue, we have been working on developing novel formulas with less cytotoxicity and superior cell internalization efficacy. For instance, we have used alkylated PEI2000 (alkyl-PEI2k) to encapsulate hydrophobic IONPs made by pyrolysis.^{22,40} Alkyl-PEI2k can hold multiple IONPs in a micelle-like



FIGURE 7. HINP-loaded macrophages were injected into a stroke model (upper right) and xenograft tumor model (lower right). Such exogenous macrophages accumulated in the areas of diseases and were detected by MRI.

nanostructure, leading to higher r₂ values and better labeling efficiency. Moreover, due to not using a long and branched PEI, the resulting nanoclusters were less cytotoxic than the previously used analogs. A second example was somewhat serendipitous, as we found that HSA-coated IONPs (HINPs) could be taken up by a wide range of cell lines at a high rate (Figure 7).^{41,42} This was unexpected, since the ζ potential of HINPs is negative (-9.46 mV), which had been thought to be suboptimal in inducing cell endocytosis. Indeed, Feridex has a ζ potential of -21.60 mV, and unless complexed with polycation material, such as PEI or poly-(L-lysine) (PLL), Feridex is insufficient to label nonphagocytic cells. One explanation to such a puzzle could be that the HSA sheath does not completely cover the intermediate dopamine coating, and the partially exposed polycation layer contributed to the cell uptake. Nonetheless, unlike the PEIcoated formulas, such HINPs have negligible cytotoxicity even at an extremely high concentration and can label a variety of cell lines without use of any excipient.

4. Magnetic Nanoparticles for Multimodality Imaging

Each imaging modality has its own advantages and disadvantages, which justifies the need for developing multimodal imaging techniques that combine the strengths of each modality and synergistically improve diagnostic quality.^{43,44} Many research activities are going on at the hardware end. For instance, SPECT/CT and PET/CT have been constructed and are being implemented worldwide. PET/MRI is under active investigation and is about to be implemented. There is, thus, a corresponding urgent need to develop multimodality imaging probes.

With a large surface-to-volume ratio and a sophisticated surface chemistry, magnetic nanoparticles can play a role as nanoplatforms, onto which non-MRI imaging motifs can be



FIGURE 8. MRI/NIRF/PET trimodal imaging (a, NIRF; b, PET; c, MRI) with HINPs that were conjugated with both ⁶⁴Cu-DOTA and Cy5.5.

easily loaded. This can immediately upgrade the agent from an MRI-only probe to an MRI-plus-X (X = PET, SPECT, NIRF, etc.) probe. For instance, we have coupled c(RGDyK) and Cy5.5 onto TPIOs. The resulting conjugates were able to home to a tumor and to depict its contour on both nearinfrared fluorescence (NIRF) and MRI images.¹⁴ Similarly, we coupled c(RGDyK) and DOTA, a macrocyclic chelator for metal binding, onto the surface of PASP–IOs.⁸ Prior to the imaging, we loaded ⁶⁴Cu, a radioisotope that is often used in PET imaging, via chelation with DOTA. The resulting nanoconjugates possessed a tumor targeting feature (due to the c(RGDyK)), as well as dual imaging capabilities via MRI (from the IONP cores) and PET (from the ⁶⁴Cu).

The advantages of such an MRI/PET or NIRF combination are substantial. The MRI can provide better anatomical information and the PET/NIRF analysis is more sensitive and quantitative or semiquantitative, allowing better assessment of the probe accumulation in the areas of interest. Such multimodality does not have to be confined to two levels. For instance, we have conjugated both ⁶⁴Cu-DOTA and Cy5.5 onto the surface of HINPs.¹¹ The resulting nanoparticles allow tumor targeting (mainly via enhanced permeability and retention) and are MRI/PET/NIRF triple functional (Figure 8). We anticipate that such a nanosystem, capable of integrating the strengths of high anatomical resolution (MRI), quantitative evaluation (PET), *ex vivo* validation (NIRF), and intraoperative potential (NIRF) will have a bright future in theranostics.

5. Magnetic Nanoparticles for Drug Delivery

There have been many efforts to use magnetic nanoparticles as vehicles for drug delivery. This immediately upgrades the nanoconjugates from MRI imaging probes to nanotheranostic agents that combine both therapeutic and diagnostic elements. Unlike other kinds of nanoparticles, such as carbon nanotubes (which are able to load therapeutics through π - π stacking⁴⁵), magnetic nanoparticles, such as

iron oxides, do not afford an easy drug loading mechanism. Until now, the drug loading on magnetic nanoparticles has been mainly on the particle coating. This again highlights the importance of surface engineering techniques.

Unlike the tethering of imaging/targeting motifs, where bioconjugation techniques are overwhelmingly used, the loading of therapeutics, although can be accomplished via covalent conjugation,^{46,47} is mostly achieved via physical means, such as electrostatic interaction. For instance, magnetic nanoparticle-based nanoplatforms have been intensively studied as gene delivery vehicles.^{47–49} The rationale is to shuttle a gene regulator (such as siRNA/shRNA/antagonist DNA) via nanoparticle vehicles across the otherwise impermeable cell membrane, where it can subsequently modulate the expression of a certain cancer-related gene. Similar to the scenario of cell labeling, nanoparticles coated with polycation materials have been widely used in such an effort. For instance, we demonstrated that alkyl-PEI2k-IOs possess many outstanding features that favor siRNA delivery, including good biocompatibility, high siRNA binding capability, protection of siRNA from enzymatic degradation, and ability to release complexed siRNA in the presence of polyanionic heparin. We observed nice gene silencing effects, at both the in vitro and in vivo levels, with siRNAloaded alkyl-PEI2k-IOs.

Magnetic nanoparticles, especially IONPs, have also been used as platforms to load small-molecule-based therapeutics. Again, since many therapeutic agents are not amenable to chemical conjugation, there is need for a nanoplatform that is able to formulate, via physical interaction, with a wide range of molecules. Lacking such an attribute themselves, magnetic nanoparticles are commonly loaded, along with therapeutics, into polymer-based matrices. More recently, we have found that albumin can be used as a good matrix material. Particularly, we found that dopamine-coated IONPs can be coloaded with therapeutics, such as paclitaxel or doxorubicin, into HSA matrices. Such a theranostic formulation takes the advantages of the well-documented, excellent ligand binding capability of HSA. Moreover, by replacing the intermediate coating layer of dopamine with caffeic acid or other dopamine analogs it is possible to tailor the surface and facilitate the loading of a broader range of therapeutics (unpublished data).

6. Conclusions and Perspectives

In summary, we and others have developed a set of chemistry to prepare magnetic nanoparticles that possess accurate sizes, shapes, compositions, magnetizations, relaxivities, and surface charges. These features, in turn, can be harnessed to adjust the toxicity and stability of the nanoparticles and, further, to load functionalities, via various mechanisms, onto the nanoparticle surfaces. These capabilities have greatly expanded the role of magnetic nanoparticles, enabling simultaneous targeting, imaging, and therapy. The close coupling of imaging and treatment within a theranostic agent and the data about the evolving course of an illness that these agents provide can facilitate informed decisions about modifications to treatment.

While the outlook is clear and exciting, it is fair to admit that we are at a relatively early stage of development. In this Account, we have dealt mostly with imaging-related, which is a true reflection of the reality. Although there is a trend toward development of more therapy-related formulas, so far this has been on the basis of chemistry and platforms that have been previously validated in the imaging setting. While multiple loading may no longer be a challenge, a more critical issue is how to leverage capabilities and to translate them into practice. The related investigations to address questions, such as how and to what extent these new formulas can advance cancer management, are being undertaken in our laboratories and those of others.

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BIOGRAPHICAL INFORMATION

Jin Xie received his Ph.D. in Chemistry from Brown University in 2008 and then joined the Molecular Imaging Program at Stanford (MIPS) as a postdoctoral research fellow. In the summer of 2009, he moved with Dr. Chen to the National Institute of Biomedical Imaging and Bioengineering (NIBIB), National Institutes of Health (NIH). His current research interests include the development and evaluation of nanoparticle- and protein-based imaging probes and drug delivery vehicles.

Gang Liu received his Ph.D. in Biomedical and Bioengineering from Sichuan University in 2009 and then joined the Laboratory of Molecular Imaging and Nanomedicine (LOMIN) of Dr. Xiaoyuan Chen at the National Institute of Biomedical Imaging and Bioengineering (NIBIB), National Institutes of Health (NIH), as a postdoctoral researcher. His current research interests focus on the development of theranostic nanomedicine systems that carry chemotherapeutics, gene therapeutics, and imaging tags.

Henry S. Eden received his M.D. from the Boston University School of Medicine in 1970 and his Ph.D. from the Department of Electrical Engineering and Computer Science of the Johns Hopkins University, Baltimore, Maryland, in 1985. He is Deputy Scientific Director, National Institute of Biomedical Imaging and Bioengineering (NIBIB), National Institutes of Health (NIH). He has a broad range of research interests at the interface of medicine and engineering.

Hua Ai received his Ph.D. in Biomedical Engineering from Louisiana Tech University in 2002. Following his doctoral studies, he completed his postdoctoral training in Biomedical Engineering at Case Western Reserve University in Cleveland, Ohio. In June 2005, Dr. Ai joined the National Engineering Research Center for Biomaterials, Sichuan University, as a Professor of Biomaterials. Currently, he also holds a joint appointment at the Department of Radiology, West China Hospital, Sichuan University. His group focuses on the design and application of self-assembly of nanobiomaterials for tumor molecular imaging and drug delivery.

Xiaoyuan Chen received his Ph.D. in chemistry from the University of Idaho in 1999. After quick postdoctoral appointments at Syracuse University and Washington University in St. Louis, he joined the University of Southern California as an Assistant Professor of Radiology. He then moved to Stanford University in 2004 and was promoted to Associate Professor in 2008. In the summer of 2009, he joined the intramural research program of the National Institute of Biomedical Imaging and Bioengineering (NIBIB) at the National Institutes of Health as a tenured senior investigator and lab chief.

FOOTNOTES

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